

**DePaul University  
Institutional Biosafety Committee  
Draft Meeting minutes  
September 17, 2025  
Location: Zoom -**

**<https://depaul.zoom.us/j/92529411190?pwd=r1TOyjI8lr4XaIzvjL4A23SwWvY2Ba.1>**

<b>Members Present</b>	Justin Maresh (Chair)	David George (Vice-Chair)	Janine Kirin
	Rima Barkauskas	Katie Abma	Jingjing Kipp
<b>Members Absent</b>	Brian Henson (alternate to Kathleen Abma)	Jeremie Fant	Nicolette Zielinski-Mozny
<b>Ex-Officio Advisors</b>	Daniela Stan Raicu*	Lauren Miller	Melodie Fox
<b>Visitors</b>			

\*Indicates not present

The September 17, 2025 meeting of the DePaul IBC for the 2025-2026 academic year began at 12:08 p.m. with a quorum present.

**I. Announcements-**

- a. Upcoming Educational Opportunities: none
- b. Upcoming IBC Meetings:
  - i. Will be scheduled shortly
- c. General Announcements:
  - i. Key Solutions will be doing an eProtocol update on 9/19/25 at 2pm which will render the system unavailable during that time. The “Cool Comments” feature will not be part of this upgrade, but other identified issues will be fixed, such as the failure to send out reminder notices for protocol expirations.
  - ii. AI Companion is being used to transcribe the audio of the IBC meeting. This transcription will not be used as Meeting Minutes; only as a supplement to the information being provided in the minutes. No objections were noted.

II. **Review of Draft Meeting Minutes**

- a. The committee members received a copy of the draft August 25, 2025 minutes prior to the meeting. No revisions were noted. Minutes were sent to the Chair for signature, which will indicate final approval.

III. **List of Meeting Minutes Approved by Chair Signature**

- a. None

IV. **New Business, Policy Discussion, and Other Discussion Items**

- i. The Chair received an email from the NIH regarding IBC policy. He will forward a copy to ORP and they will discuss the dissemination of information to the rest of the IBC.
- ii. The IBC Policy Manual will be reviewed by September 19, 2025 to confirm whether or not meeting minutes need to be voted on.

V. **Continuing Education Materials and Discussion**

- a. None

VI. **Current Protocol Submissions for Review**

- a. Renewals: none
- b. Amendments: none
- c. New Protocols that the IBC requested modifications to be brought to the full board: none
- d. New Protocols for Initial Review:
  - i. Research Protocols
    - a. Protocol #: 2025-1715  
Title: The Biology of *Stenotrophomonas maltophilia*  
PI Name: Dr. Joanna Brooke  
Biohazardous Agent(s): *Stenotrophomonas maltophilia* (*S. maltophilia*), *Pseudomonas aeruginosa* (*P. aeruginosa*), *Escherichia coli* (*E. coli*), *Staphylococcus aureus* (*S. aureus*), *Staphylococcus epidermidis* (*S. epidermidis*), EZ-Tn5™ <R6Kγori/KAN-2>Tnp Transposome™, Phages, *Enterococcus faecalis* (*E. faecalis*), Miscellaneous bacterium  
RG: 1 & 2  
BSL: 1 & 2  
NIH category: III-D-2-a, III-F-3  
Primary Reviewers: David George and Justin Maresh

**Protocol Summary:** The objective of Project 1 is to understand how the emerging multi-drug resistant opportunistic pathogen, *Stenotrophomonas maltophilia*, can result in serious infections in immunocompromised patient populations. This ubiquitous

bacterium can form bacterial films (biofilms) on the surfaces of medical equipment in hospitals and implanted devices in patients. These biofilms can act as pervasive and persistent sources of nosocomial infections by *S. maltophilia*. These observations underscore the need to understand the molecular mechanisms involved in the biology of this pathogen and develop new strategies against *S. maltophilia*. Using cell based assays, biochemical methodologies, molecular biology techniques and microscopy, we intend to study mutant and wild type strains of *S. maltophilia*, their cell structure, genetics and their ability to survive and persist when challenged with antimicrobials and select chemicals. Project 2 will investigate bacteriophages (phages) and how they interact with *S. maltophilia* in the environment. This project will use *S. maltophilia* strains and mutants to recover phages from environmental samples. Phages will be enriched and isolated from these samples using standard collection techniques. Cell assays, microscopy, microbiology, and molecular biology techniques will be used to identify and characterize the phages, and assess their interaction with *S. maltophilia* and its biofilms under different culture conditions. Project 3 will isolate *S. maltophilia* from freshwater isopods (invertebrates). The ability of the isolated *S. maltophilia* to grow and form biofilms in different culture conditions will be tested and the bacterium's resistance to antimicrobials will be studied. Project 4 will isolate, identify and characterize bacteria that have been recovered from surfaces. Bacteria will be tracked to serve as an indicator of how much microbial contamination is present. The survival, growth and biofilm of these bacteria in different culture conditions will be studied.

### **Discussion:**

At the September 17 meeting, the IBC discussed the following issues:

- Maria Robledo needs to take the CITI IBC training.
- Timothy Sparkes is on leave, but it is unclear for how long. Before working on this protocol, he needs to take the Lab Safety Training.
- *Pseudomonas aeruginosa* (*P. aeruginosa*) is listed in the biohazardous agents table, however it is not included in the project. If it is not to be included in the project, it must be deleted from the table. If it is to be included in the project, the project information needs to include when and how it is to be used.

- The biological safety cabinet certification dates need to include the month and year.
- There is a discrepancy between the rooms listed for sanitation and disposal and the ones listed in the SOP. Room 127 needs to be added to the protocol for waste collection and room 204 needs to be added to the SOP.

### **IBC Determinations:**

- The agent(s) meets the criteria for Risk Group (RG) 1 and 2.
- This protocol requires biosafety containment level 1 and 2 procedures under the *NIH Guidelines* or the *BMBL* for the biohazardous agent being utilized.
- This protocol continues to falls under NIH Category(ies) III-D-2-a, III-F-3
- Modifications requested to be reviewed by the primary reviewers.

A motion was made and seconded to request modifications for Protocol #2025-1715. The vote was as follows:

Motion: Modifications to be Required to be Reviewed via DMR	For: 5	Against: 0	Abstain: 0	Recused: 0	Total: 5
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#### *b. Protocol #: 2025-1727*

Title: PCB Effects on Microglia in vitro

PI Name: Margaret Bell

Biohazardous Agent(s): Lipopolysaccharide, LPS, derived from *E. coli* 0111:B4; Polychlorinated biphenyls (PCBs) in Aroclors A1242, A1248, and A1254, including PCB 153 and 28; Polychlorinated biphenyls (PCBs) and LPS diluted in cell culture media

RG: 1 & 2

BSL: 1 & 2

NIH category: N/A

Primary Reviewers: Justin Mares & Katie Abma

**Protocol Summary:** This proposal seeks to understand if and how exposure to an environmental contaminant (polychlorinated biphenyls, PCBs) affects inflammatory processes in the brain. A primary culture of brain cells will be exposed to PCBs and challenged with an inflammogen, lipopolysaccharide, LPS, essentially 'faking cells into thinking they are sick'. Protein or RNA will be collected from cells in the hours to days that follow and quantify changes in cell activity and number. The main goals are to determine if PCB exposure alters the normal and activated immune

processes in the brain and identify specific proteins and cells that are part of this process.

To test the interactions between developmental PCB exposure and neuroinflammation, primary microglial culture will be used to assess effects of PCBs on the cells directly. Cultures will be treated with an low concentration solution of PCBs (0.1 - 10 micromol) in DMSO vehicle. 1-24 hours later, cells will be treated with lipopolysaccharide (LPS, derived from *E. coli*, 1 ug/ml). After PCB and LPS treatments, cells will be collected and lyzed to isolate RNA, and culture media will be collected to isolate protein. Changes in gene expression and protein production will be quantified using previously established protocols. Genes and proteins for analysis will be identified based on scientific literature documenting key PCB targets and inflammation processes.

### **Discussion:**

At the September 17 meeting, the IBC discussed the following issues:

- Julie Pavlova needs to take the IBC CITI training entitled Faculty/Teaching Assistants/Lab Coordinators/Research Personnel (including Grad Students)
- Safety Glasses/goggles needs to be checked off since they are mentioned being used.
- Duplicate copies of SOPs were attached. The newer versions were reviewed by EHS. The DePaul Student Health information was correct in one version but not the other. All mentions of Sage Medical need to be removed. The protocol numbers on the SOPs also need updating.
- Cultured rat cells need to be included in the biohazardous agent table as RG1, BSL1 due to their being a potential risk of unknown biohazard.

### **IBC Determinations:**

- The agent(s) meets the criteria for Risk Group (RG) 1 and 2.
- This protocol requires biosafety containment level 1 and 2 procedures under the *NIH Guidelines* or the *BMBL* for the biohazardous agent being utilized .
- Modifications requested to be reviewed by the primary reviewers.

A motion was made and seconded to request modifications for Protocol #2025-1727. The vote was as follows:

Motion: Modifications to be	For: 5	Against: 0	Abstain: 0	Recused: 0	Total: 5
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Required to be Reviewed via DMR					
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ii. Teaching Protocols: none

VII. **Notification of Approval Actions for Protocols Submissions in which the IBC Requested Modifications:** none

VIII. **Notification of Approval Actions Conducted Under Designated Member Review:**

a. Renewals: none

b. Amendments: none

IX. **Notification of Approval Actions Conducted Under Administrative Review:** none

X. **Notification of Protocols Confirmed at Exempt via Chair Confirmation:** none

XI. **Notification of Protocols that Have Been Terminated or Suspended:** none

XII. **Notification of Protocols that Have Been Administratively Closed:** none

The meeting ended at 12:45 p.m.

Submitted respectfully by,

Janine Kirin  
Director of Research Support Facility  
On behalf of the Office of Research Services

Approved by IBC Chair Signature:



Justin Maresh, Ph.D.  
IBC Chair

Date: 11/3/2025